

Gene Therapy for Haemophilia

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Declaration of financial interests by Edward Tuddenham

- I am a named inventor of patents in the field of gene therapy for haemophilia held by UCL
- I am a consultant to Biomarin Inc.
- I am a consultant to Freeline Therapeutic Ltd.

AAV Transduction Pathway

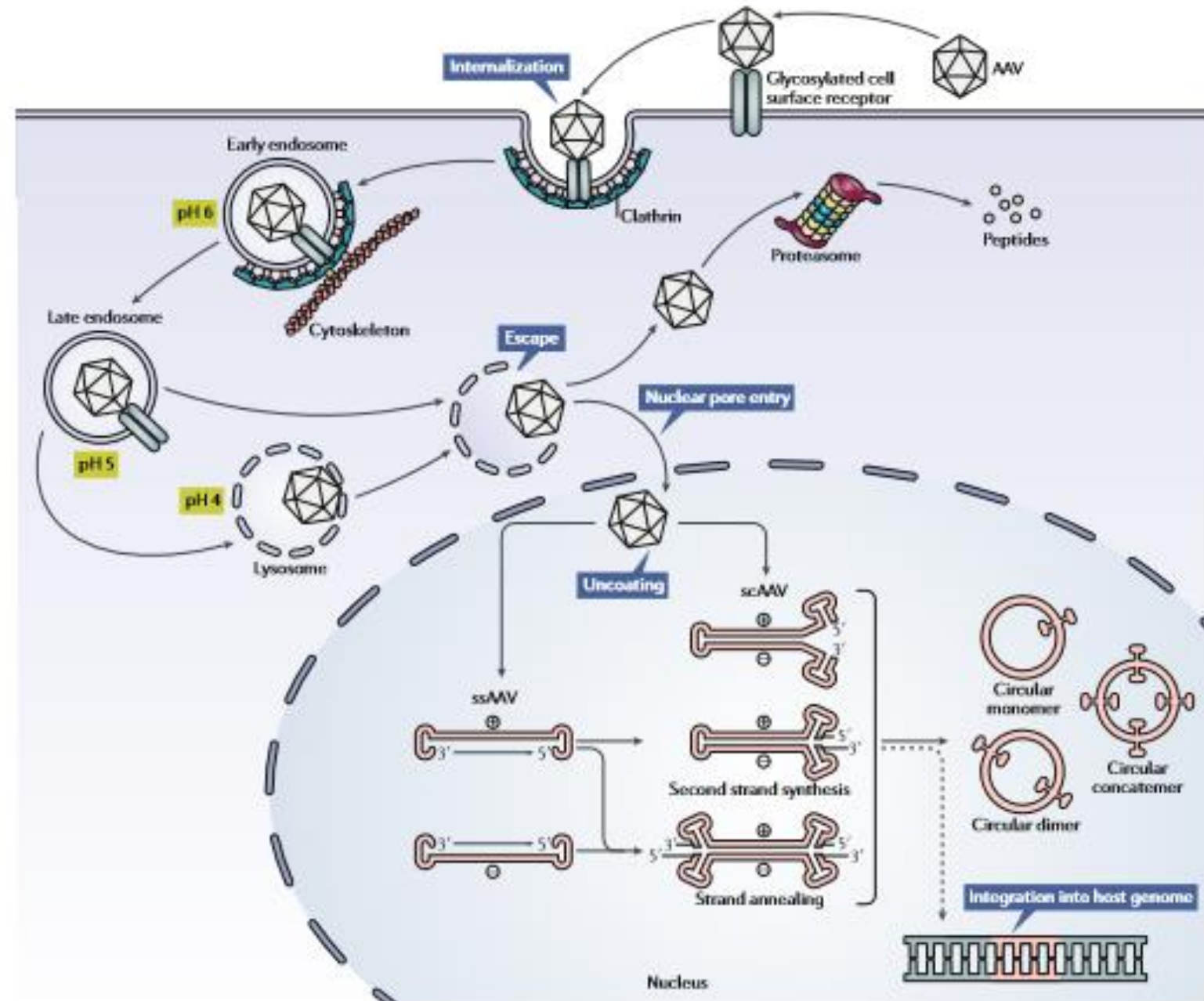
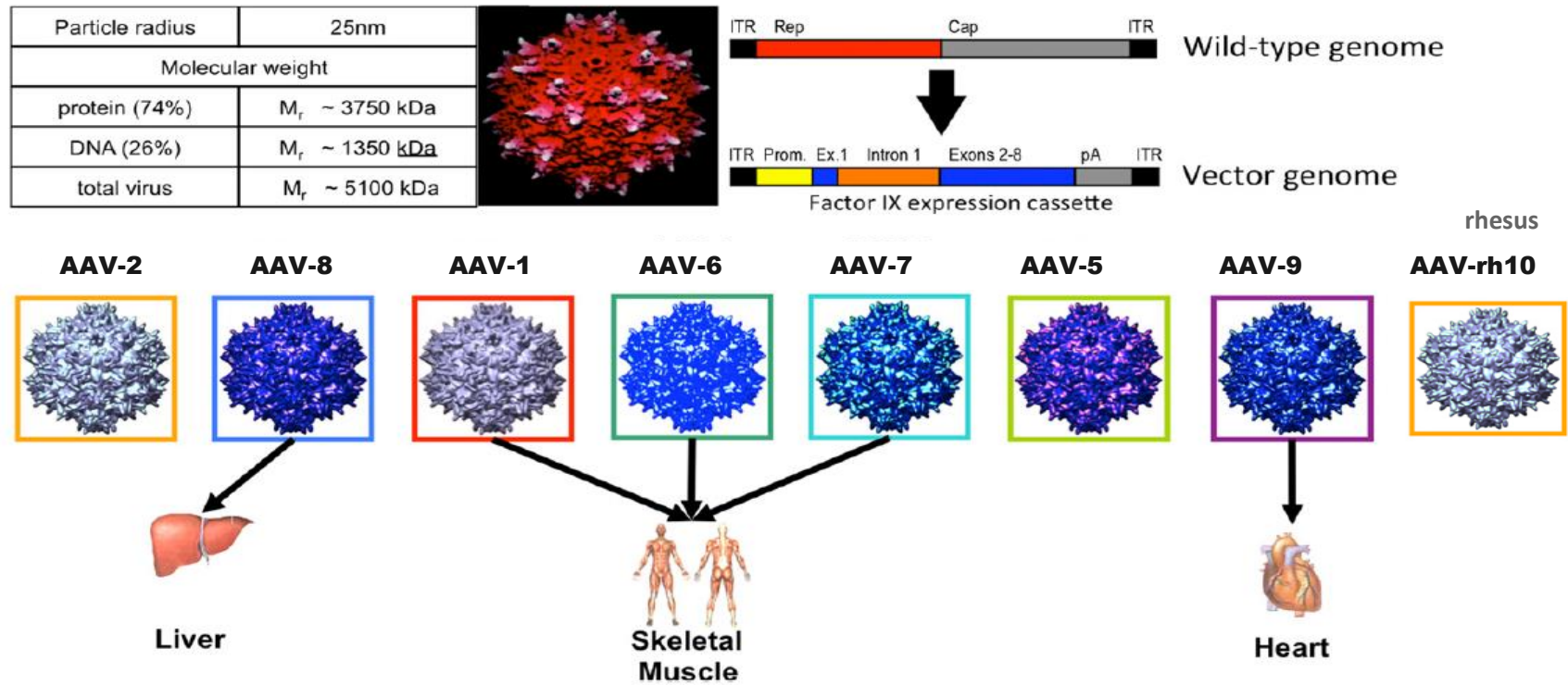


Fig. 2 | **Diagram of rAAV transduction pathway.** Adeno-associated virus (AAV) is recognized by glycosylated cell surface receptors of the host. This triggers internalization of the virus via clathrin-mediated endocytosis. AAV then traffics

AAV Vectors: Gene Transfer



Spark SPK 8011
AAVspk FVIII

Sangamo SB-525
AAV6 FVIII

BioMarin BMN 270
AAV5-FVIIIISQ

Dimension DTX201
AAV rh10 FVIII

UCL/St Jude
AAV8-FVIIIv3

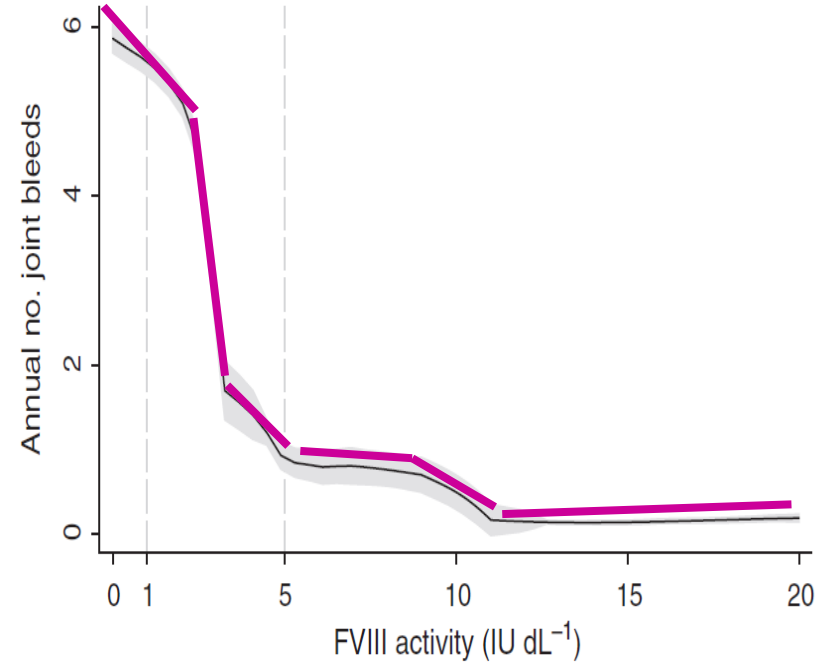
Shire SHP654
AAV8 FVIII

Rationale: Gene Therapy for Hemophilia

- Hemophilia is monogenic
- Wide range of FVIII, IX effective
- Tissue-specific expression not required
- Well-characterized animal models exist
- One-time treatment required for gene transfer
- Avoids cost, morbidity, complications of current therapy
- Potential global treatment for all affected

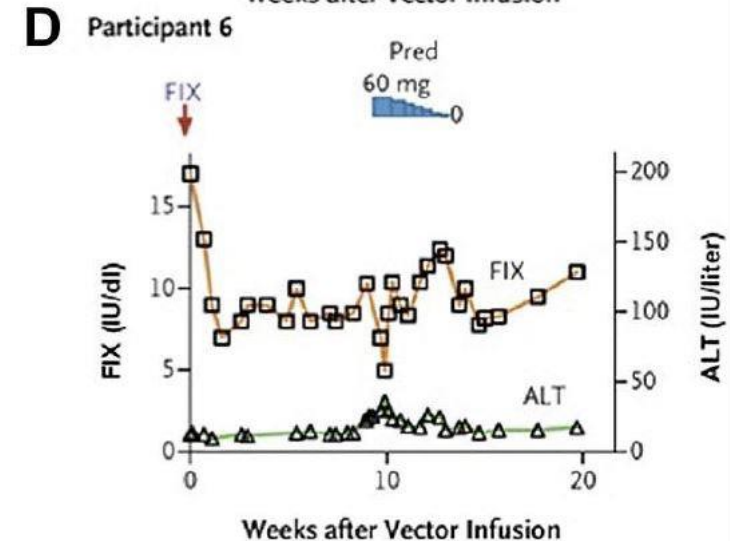
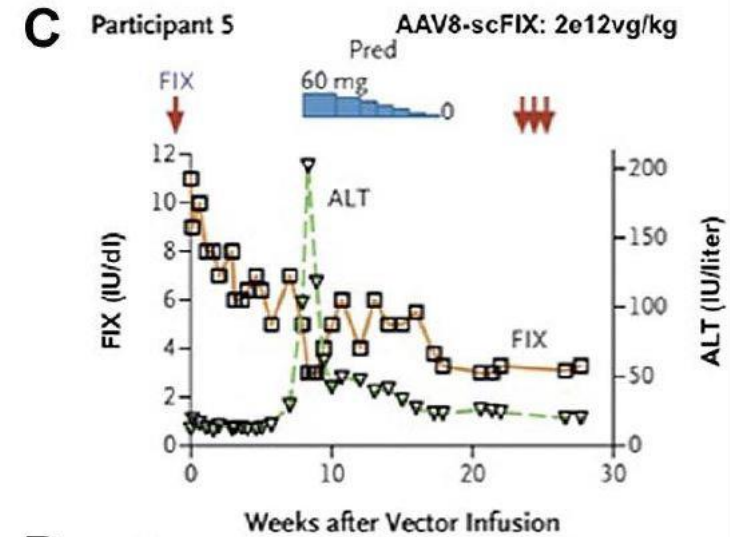
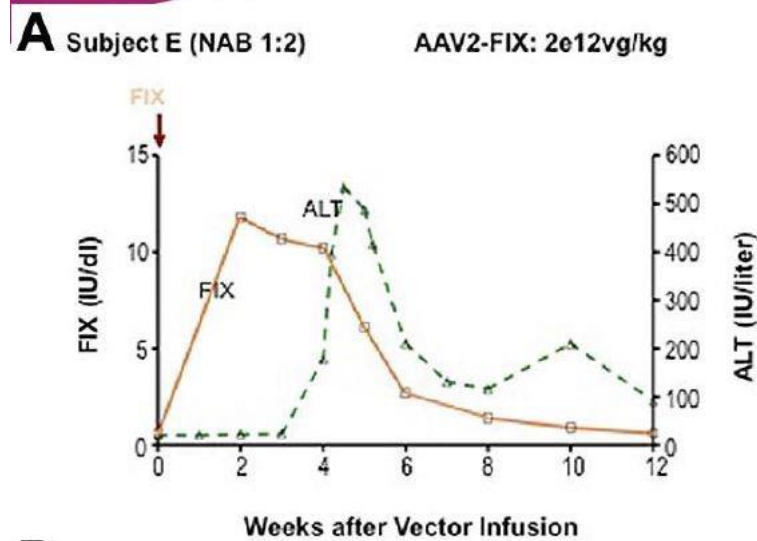
Aim: Hemophilia Gene Transfer

- **Sustain:** long-term expression
- **Achieve:**
 - Past:* FVIII, IX $\geq 15\%$
 - Current:* FVIII, IX $> 50\%$
- **Convert:** severe to normal
- **Avoid:** all bleeds



Two trials that established gene therapy for haemophilia B

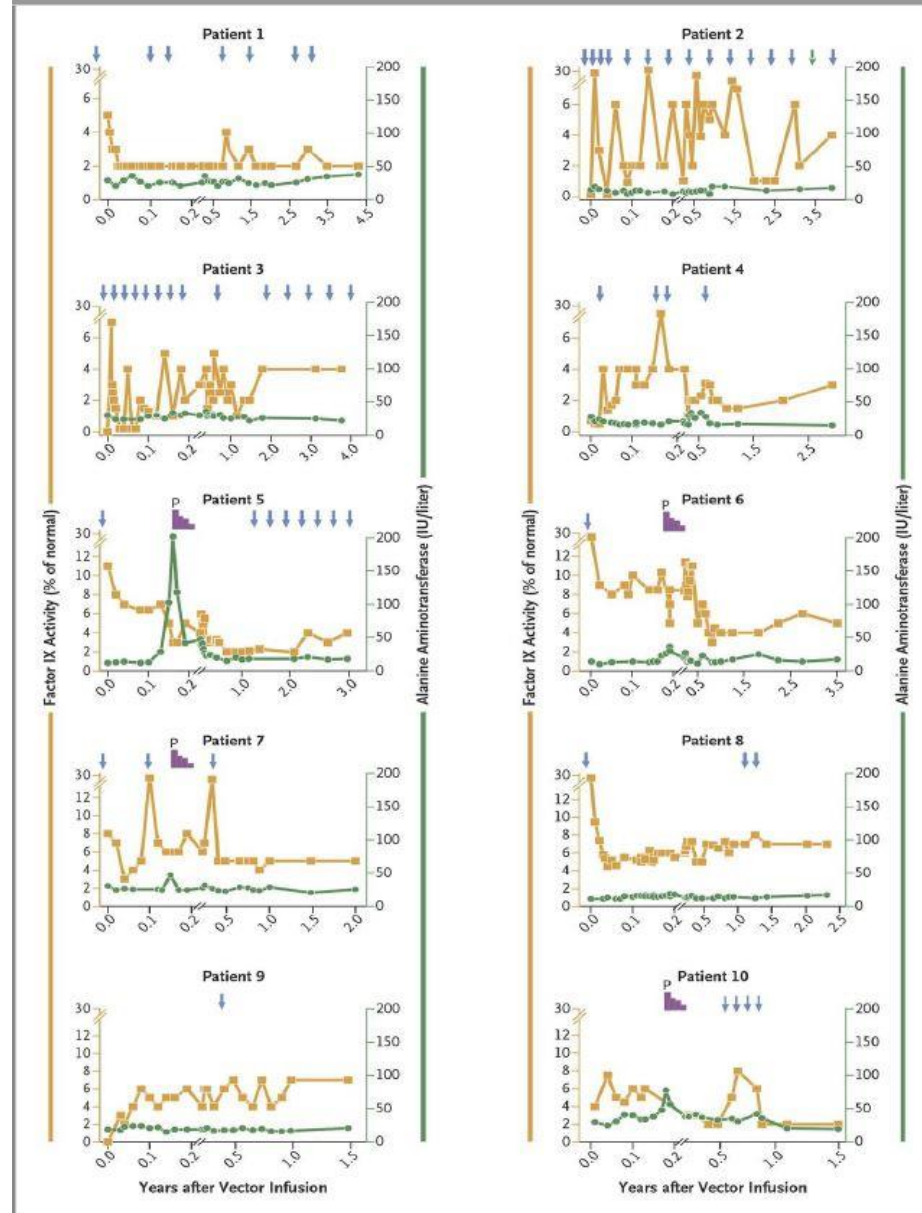
Manno et al 2006; Nathwani et al 2010



Successful transduction of liver in hemophilia by AAV-Factor IX and limitations
e. *Nat Med.* 2006;12(3): 342-347.

Nathwani S, et al. Adenovirus-associated virus vector-mediated gene transfer in
(25):2357-2365.

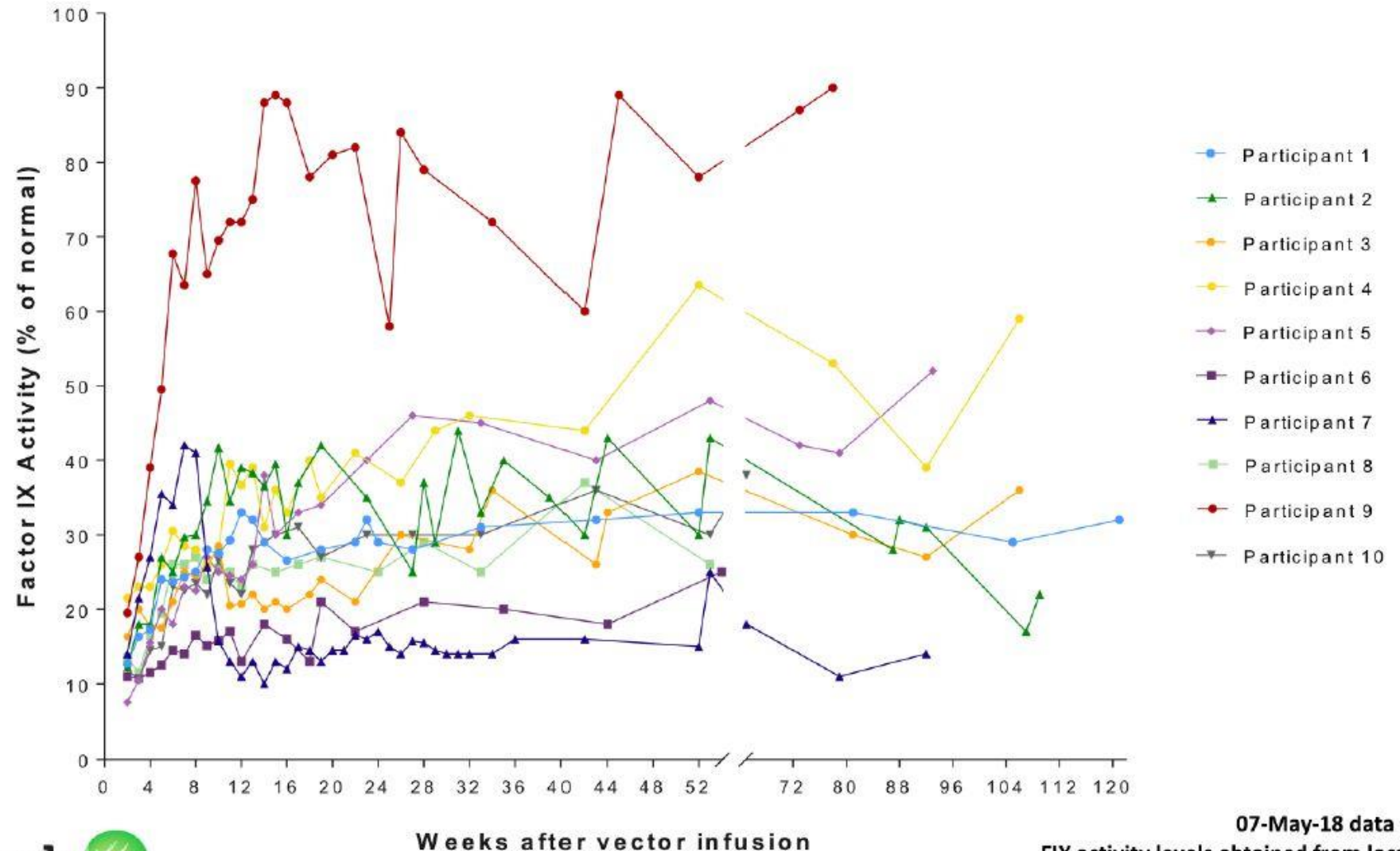
Longer term follow up published in 2014 is considered a 'landmark study'



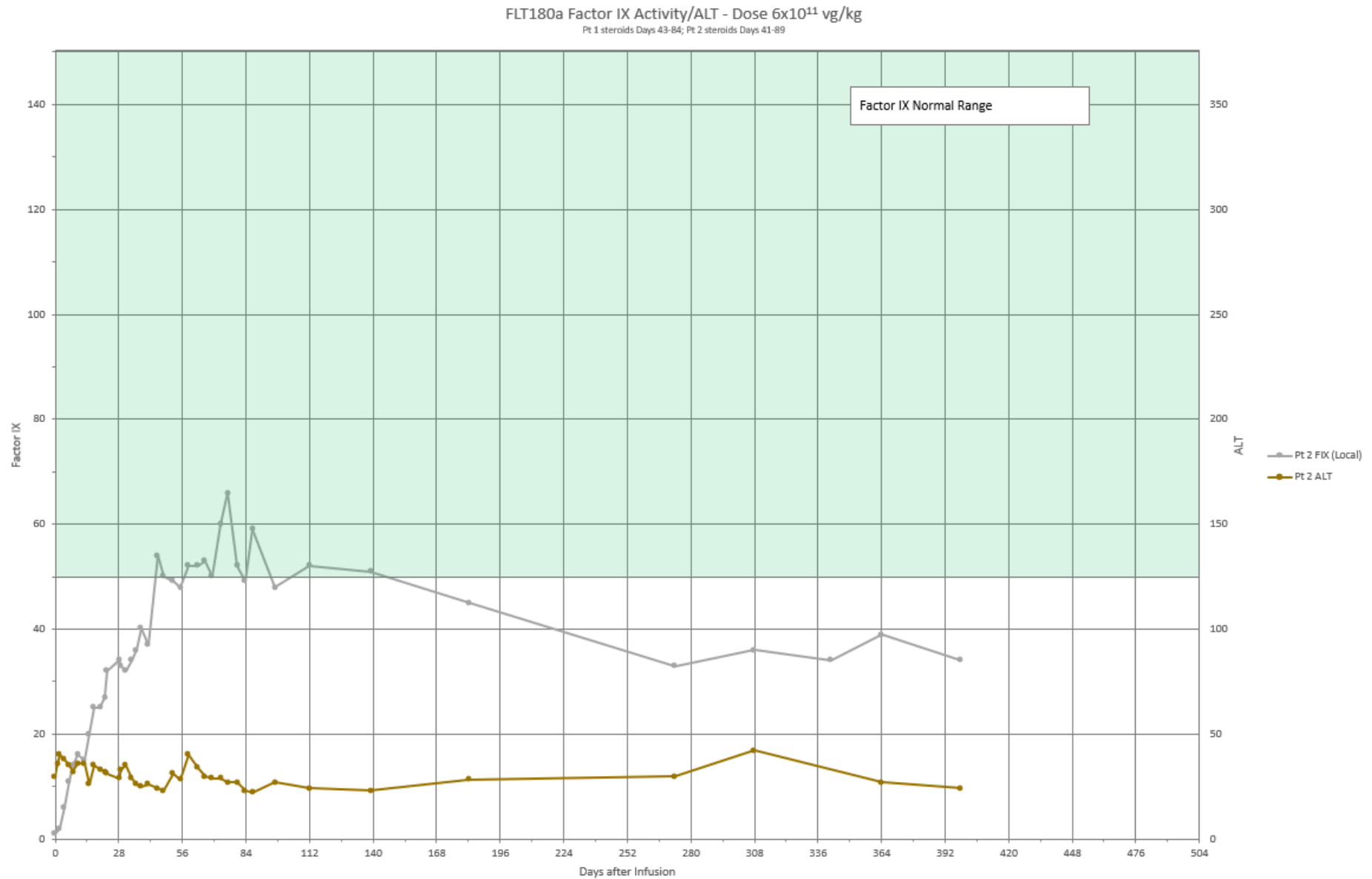
1. Data for up to 4 years
2. Consistent and durable response
3. No lasting adverse effects
4. Liver enzyme elevation controlled by steroid treatment
5. Bleeding rate reduced substantially
6. Factor usage reduced to near zero in 6 of 10 patients.

New study in haemophilia B using high activity factor IX Padua variant

- Pioneered successful trial with R338L FIX
- Range of mean steady-state FIX activity: 14.3-76.8%
- 2 transient transaminase elevations
- 3 received steroids
- Evidence of anti-capsid response
- No inhibitors
- Profound reduction in bleeding rate and factor utilization through 2 years



Freeline study with Padua Factor IX in rAAVS3 capsid dose 6×10^{11} vg/kg



A possible cause of “transaminitis”

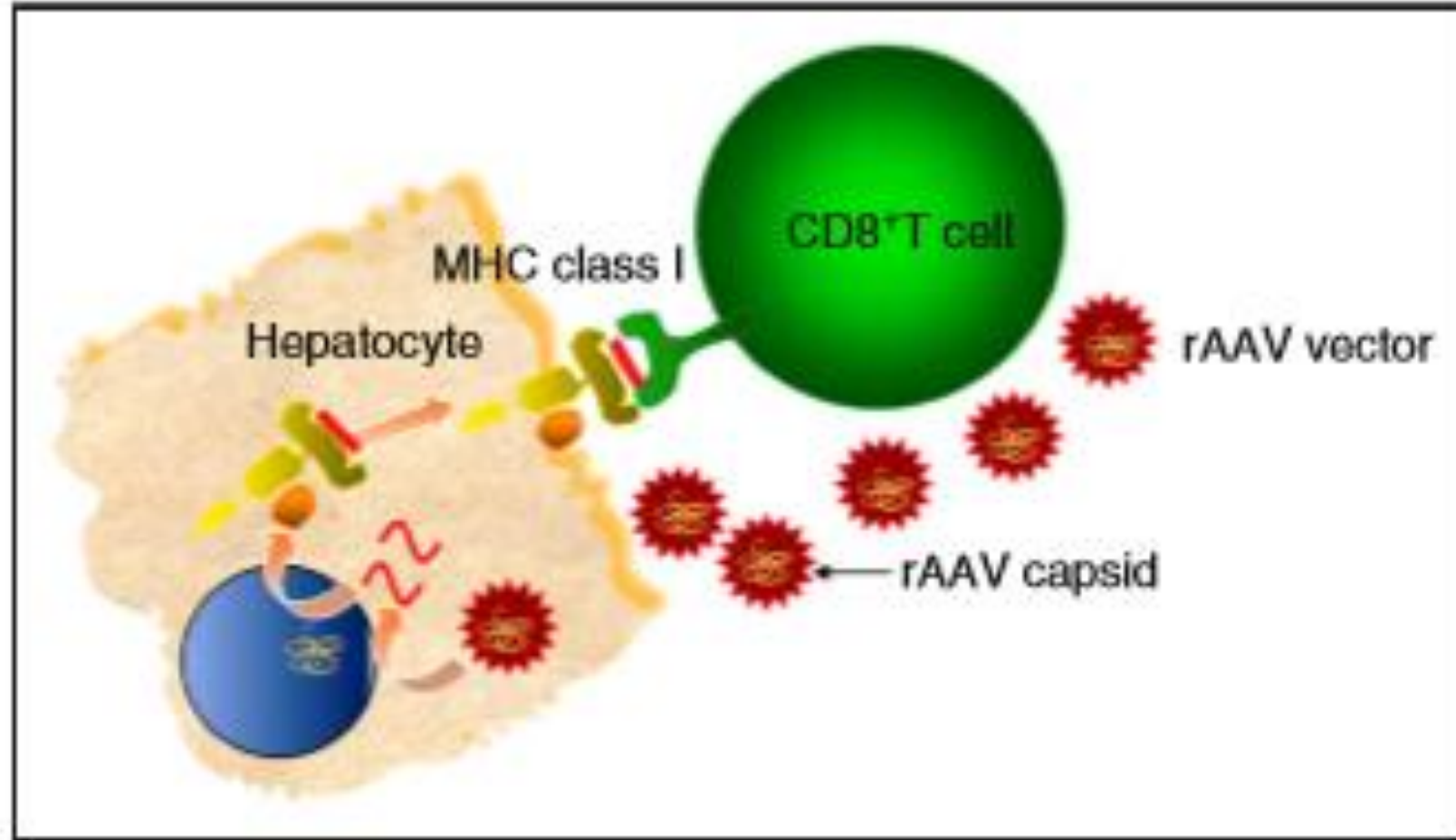


Figure 1. Hypothesis of the cellular immune response to hepatocytes after rAAV transduction. rAAV enters the hepatocyte by receptor-mediated endocytosis,

Immune barriers to successful transduction by AAV vectors in humans

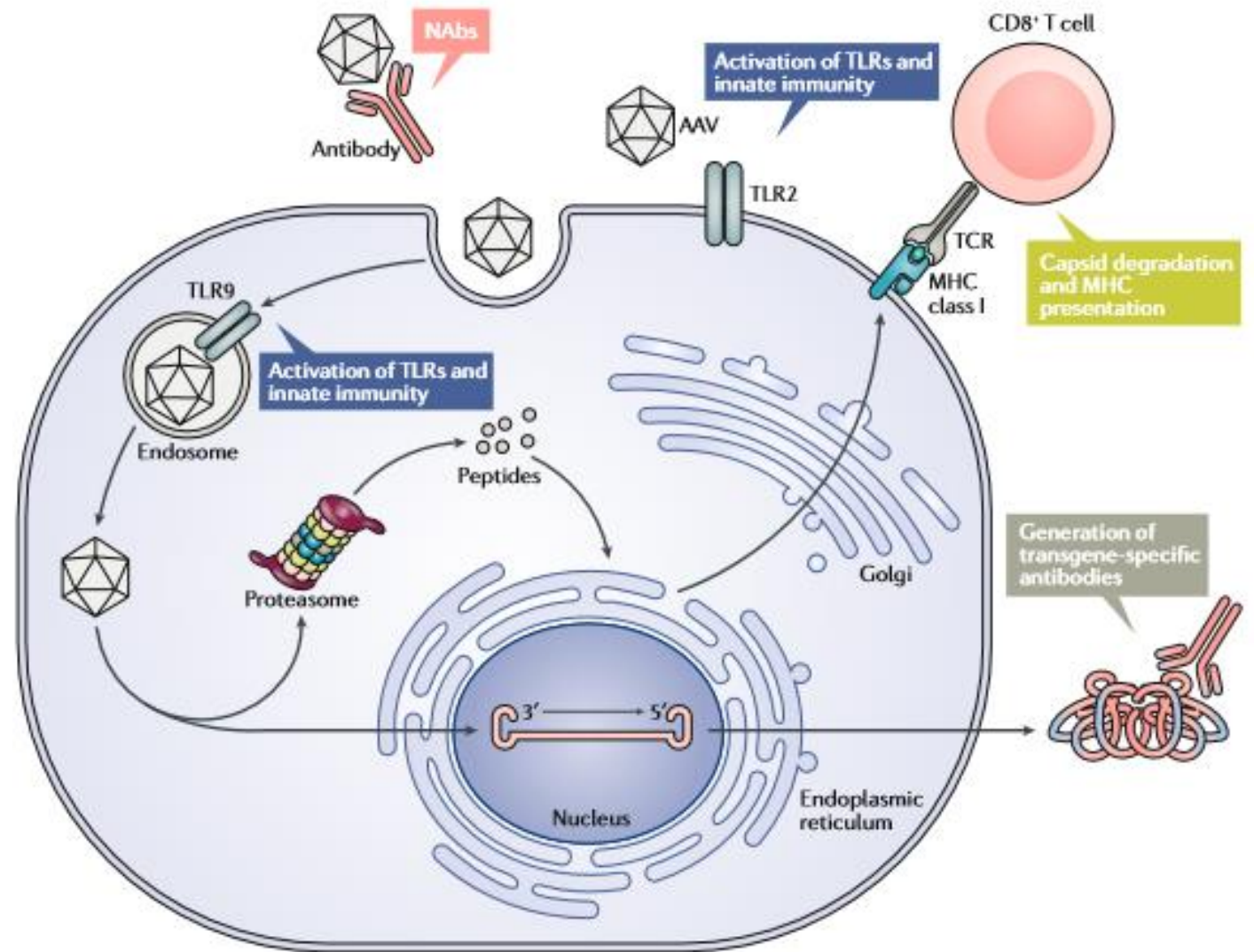


Fig. 5 | **Immunological barriers to successful rAAV gene delivery.** The recombinant adeno-associated virus (rAAV) may

Rapid increase of investment and new trials using AAV 2014-2019

Table 1 | A selection of ongoing rAAV interventional clinical trials

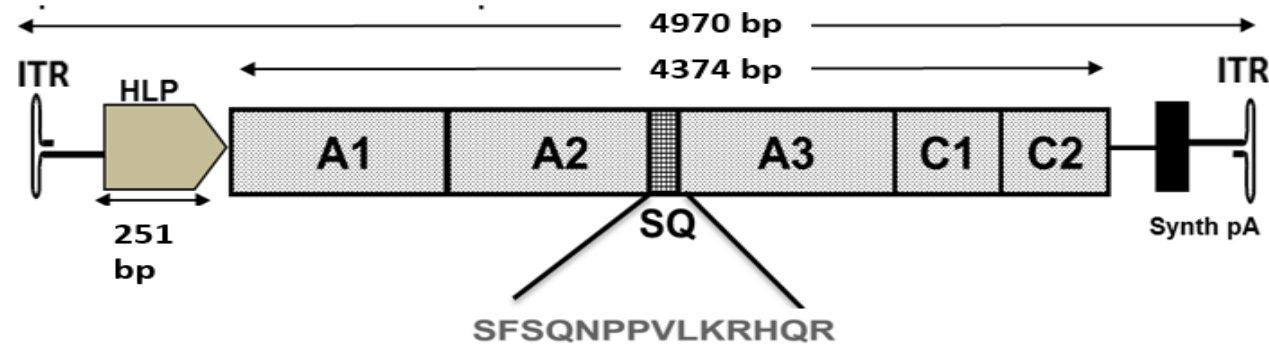
Primary gene delivery target	Condition	AAV capsid	Transgene product	Strategy	Sponsor	Phase	ClinicalTrials.gov identifier
Brain	AADC deficiency	AAV2	AADC	Replacement	Kryofa/BioKryofa, UCSF	Phase I	NCT01852713
		AAV2	AADC	Replacement	National Taiwan University Hospital	Phase II	NCT02929866
	Batten disease (CIN2)	AAVrh.10	CIN2	Replacement	Welli Cornell	Phase I/II	NCT01404905
	Batten disease (CIN6)	AAV9	CIN6	Replacement	Nationwide Children's Hospital	Phase I/II	NCT02725580
	MPS-III	AAV5	NAGLU	Replacement	uniQure	Phase I/II	NCT01300453
	Parkinson disease	AAV2	AADC	Addition	Jichi Medical University	Phase I/II	NCT02408598
		AAV2	GDNF	Addition	MINDS	Phase I	NCT01621581
Spinal cord	SMA	AAV9	SMN	Replacement	AveXis	Phase II	NCT03481289
		AAV9	GAN	Replacement	MINDS	Phase I	NCT03382438
Eye	Achromatopsia	AAV2	CNGH3	Replacement	AGTC	Phase I/II	NCT02549902
		AAV8	CNGH3	Replacement	MeiraGTX	Phase I/II	NCT03081310
	Choroideremia	AAV2	REP1	Replacement	Nightstar	Phase II	NCT03496012
		AAV2	REP1	Replacement	Spark	Phase I/II	NCT02340807
		AAV2	REP1	Replacement	SEZ ocular	Phase II	NCT02671539
		AAV2	REP1	Replacement	University of Oxford	Phase II	NCT02402578
	LCA	AAV2	RPE65	Replacement	Spark	Phase II	NCT00999609
		AAV5	RPE65	Replacement	MeiraGTX	Phase I/II	NCT02381480
	LHON	AAV2	ND4	Replacement	GenSight	Phase II	NCT02915254
		AAV2	ND4	Replacement	Johns Hopkins University of Miami	Phase I	NCT02351380
	RP (RBP1)	AAV8	RBP1	Replacement	Novartis	Phase I/II	NCT03324857
	Wet AMD	AAV8	Anti-VEGF antibody	Silencing (mAb)	Regeneron	Phase I	NCT03066258
	X-linked RP	AAV2	RPCR	Replacement	AGTC	Phase I/II	NCT03338560
		AAV2	RPCR	Replacement	MeiraGTX	Phase I/II	NCT03252947
		ND	RPCR	Replacement	Nightstar	Phase I/II	NCT03138115
	X-linked retinoschisis	AAV2	RS1	Replacement	AGTC	Phase I/II	NCT02436632
		AAV8	RS1	Replacement	NEI	Phase I/II	NCT02317887
Liver	Crigler-Najjar syndrome	AAV8	UGT1A1	Replacement	Audentes	Phase I/II	NCT02723194
		ND	UGT1A1	Replacement	Genethon	Phase I/II	NCT03466493
	EH (hemochromatosis)	AAV8	EDR	Replacement	University of Pennsylvania	Phase I/II	NCT02655675
	GSD1a	AAV8	G6PC	Replacement	Ultragenyx	Phase I/II	NCT03517085
		AAV8	FVIII	Replacement	Shire	Phase I/II	NCT03108172
		AAVrh.37	FVIII	Replacement	Royce	Phase I/II	NCT03588299
		AAV5	FVIII	Replacement	BioMarin	Phase II	NCT03162974
		AAV5	FVIII	Replacement	Sangamo	Phase I/II	NCT03061201
		ND	FVIII	Replacement	Spark	Phase I/II	NCT03007533
	Haemophilia A	AAV8	FVIII	Replacement	UCL	Phase I	NCT03008330

Table 1 (cont.) | A selection of ongoing rAAV interventional clinical trials

Primary gene delivery target	Condition	AAV capsid	Transgene product	Strategy	Sponsor	Phase	ClinicalTrials.gov identifier
Liver	Haemophilia B	AAV8	FIX	Replacement	Shire	Phase I/II	NCT01687608
		ND	FIX	Replacement	Pfizer	Phase II	NCT02484092
		ND	FIX	Replacement	Pfizer	Phase III	NCT03587116
		AAV6	FIX	Replacement	Sangamo	Phase I	NCT02695160
		AAV8	FIX	Replacement	St. Jude Children's Research Hospital	Phase I	NCT00979238
		AAV5	FIX	Replacement	uniQure	Phase III	NCT03569891
	MPS-I	ND	FIX	Replacement	UCL	Phase I	NCT03369444
		AAV6	ZFN1, ZFN2 and IDUA donor	Editing	Sangamo	Phase I	NCT02702115
	MPS-II	AAV6	ZFN1, ZFN2 and IDS donor	Editing	Sangamo	Phase I	NCT03041324
	MPS-III	AAVrh.10	SGSH	Replacement	LYSOGENE	Phase II/III	NCT03612869
Muscle	MPS-VI	AAV8	ARSB	Replacement	Fondazione Telethon	Phase I/II	NCT03173521
	OTC deficiency	AAV8	OTC	Replacement	Ultragenyx	Phase I/II	NCT02991144
	A1AT deficiency	AAV2	A1AT	Replacement	UMMS	Phase I	NCT00377416
		AAV1	NTF3	Addition	Nationwide Children's Hospital	Phase I/II	NCT03520751
	DMD	AAVrh.74	Micro-dystrophin	Replacement	Nationwide Children's Hospital	Phase I/II	NCT03375164
		AAV9	Mini-dystrophin	Replacement	Pfizer	Phase I	NCT03362502
		AAV9	Micro-dystrophin	Replacement	Solid Biosciences	Phase I/II	NCT03368742
	Dysferlinopathy	AAVrh.74	DYSF	Replacement	Nationwide Children's Hospital	Phase I	NCT02710500
	HIV infections	AAV1	PG9 antibody	Addition	International AIDS Vaccine Initiative	Phase I	NCT01937455
		AAV8	VRC07 antibody	Addition	NIAID	Phase I	NCT03374202
Pompe disease	Pompe disease	AAV8	GAA	Replacement	Actus Therapeutics	Phase I/II	NCT03533673
		AAV9	GAA	Replacement	University of Florida	Phase I	NCT02240407
	X-linked MTM	AAV8	MTM1	Replacement	Audentes	Phase I/II	NCT03199469

A1AT, α1-antitrypsin; AADC, aromatic L-amino acid decarboxylase; AGTC, Applied Genetic Technologies Corporation; AMD, age-related macular degeneration; AAV8, serotype B; CIN2, neuronal ceroid lipofuscinosis type 2; CMT1A, Charcot-Marie-Tooth disease type 1A; CNGH3, cyclic nucleotide-gated channel, β2;

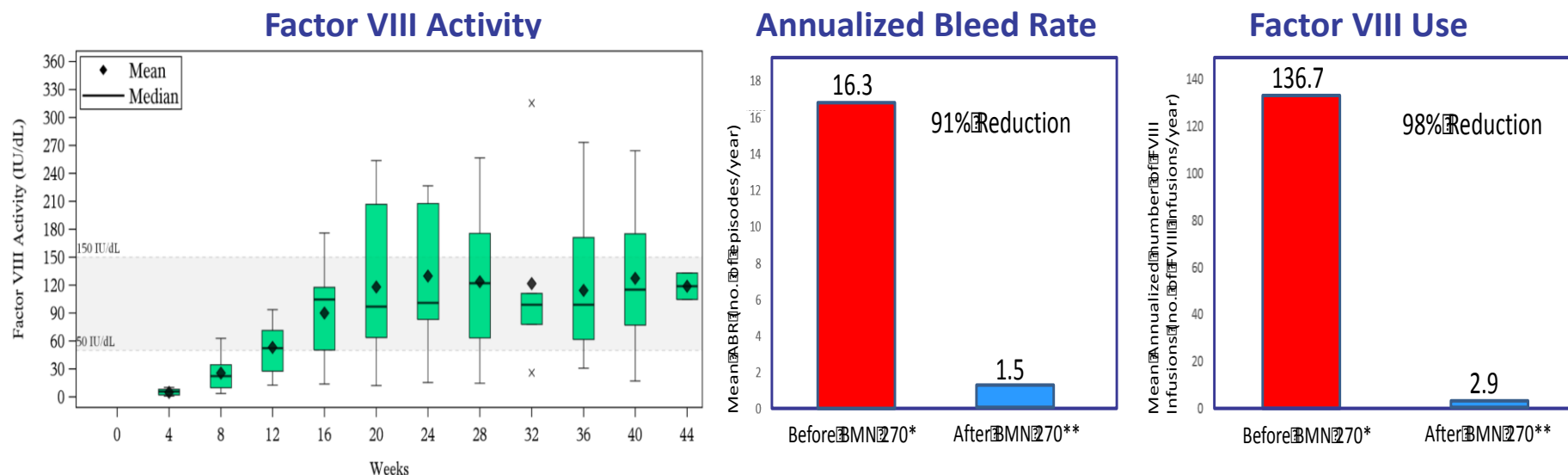
BMN 270: AAV vector construct



- Due to packaging constraints, size of vector components minimized
- Efficient transduction of the liver by AAV5
- Liver specific promoter
- Active Factor VIII with B domain deletion of Refacto™

AAV5-hFVIII

Biomarin: BMN270 – AAV5-BDD-FVIII



Clinical Findings:

- **Factor VIII Activity:** Mean peak 115-130%
- **Durability:** Maintained to > 44 weeks
- **ABR:** 91% reduction (No bleeds in 5/6)
- **FVIII Use:** 98% reduction (No FVIII in 5/6)
- **Immune response:** High-dose cohort - steroid responsive

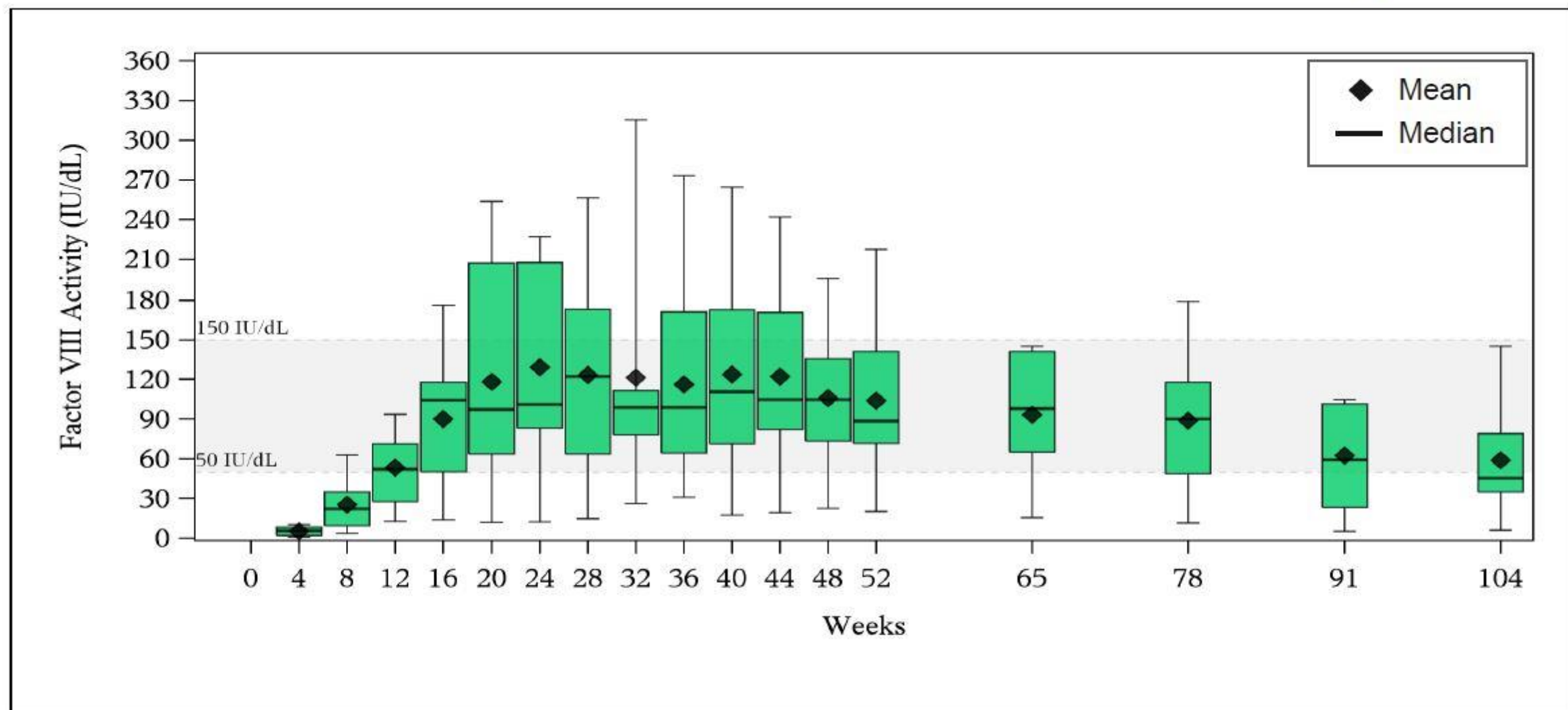
Conclusion: Safe, effective, durable > 44 weeks
Steroid-responsive immune response

Biomarin, 2016, 2017

Liver Function Tests

	Peak ALT	Last ALT	Status
1	60	15	Normal
2	95	16	Normal
3	82	42	Normal
4	87	33	Normal
5	43	38	Normal
6	81	45	<1.1 ULN
7	66	27	Normal

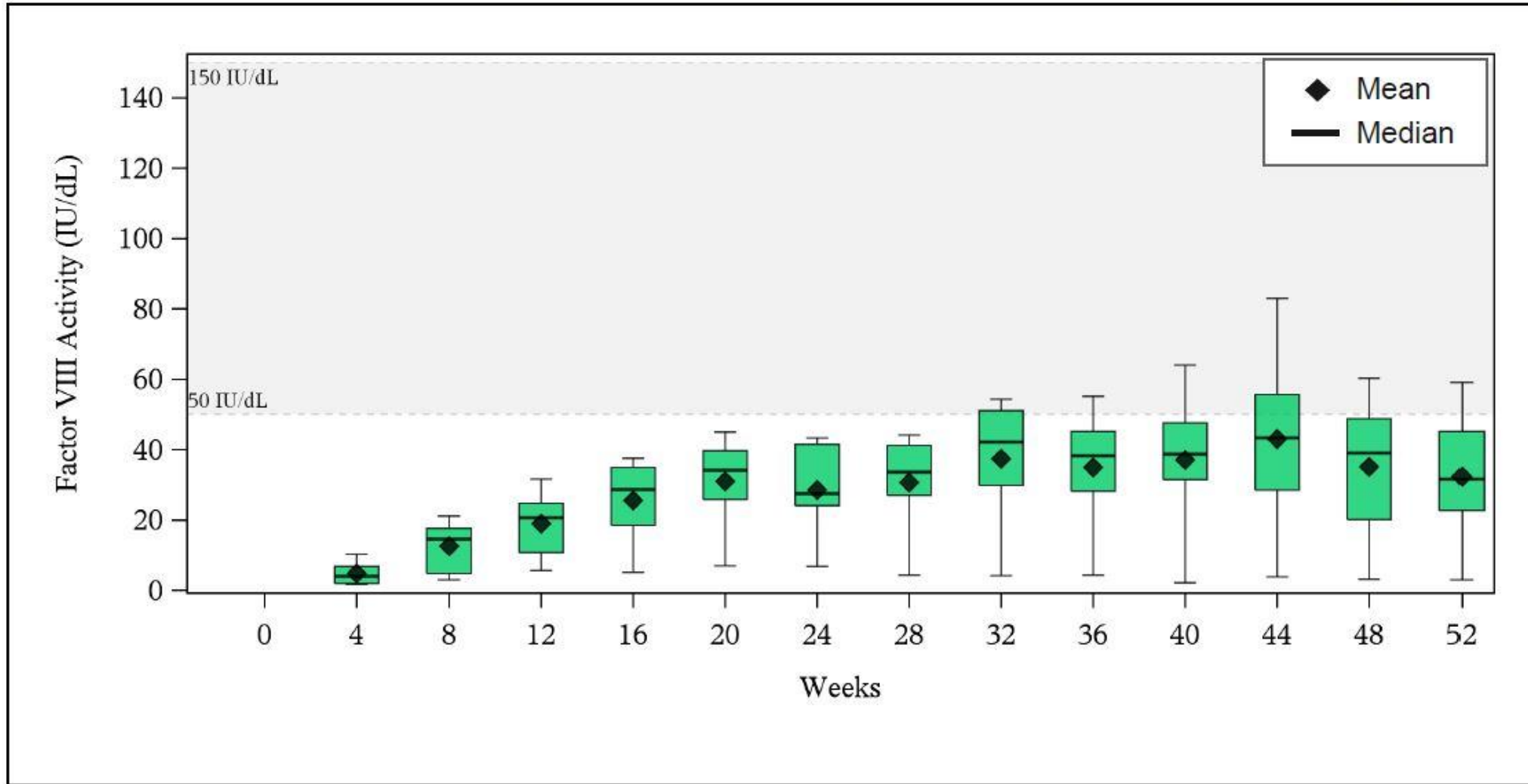
MEAN FVIII ACTIVITY LEVELS SETTLING IN NORMAL RANGE (6e13 VG/KG)



- No FVIII activity above upper limit of normal at year 2

The upper and lower box bounds represent 25th and 75th percentiles. The whisker lines represent the minimum and maximum values.

MEAN FVIII ACTIVITY LEVELS AT HIGH END OF MILD RANGE (4e13 VG/KG)



- No FVIII activity above normal

3 year data from BioMarin study

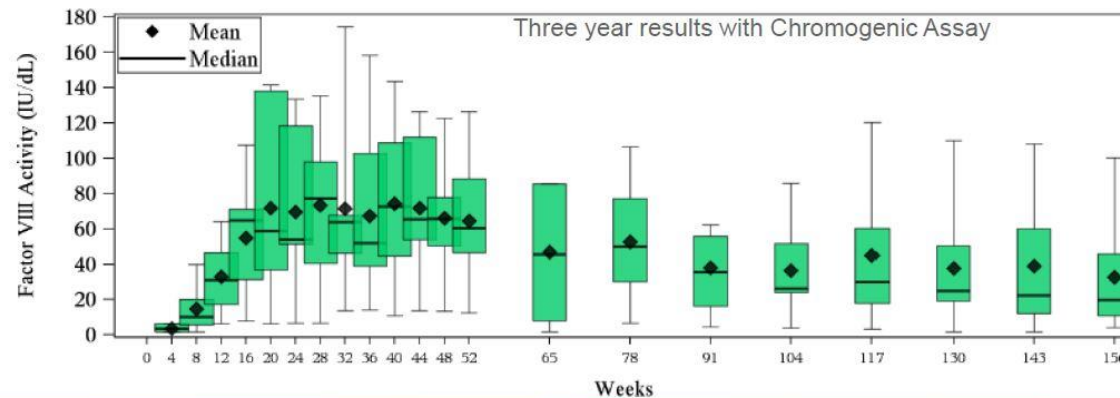
BiOMARIN

Valoctocogene Roxaparvovec Phase 2 results

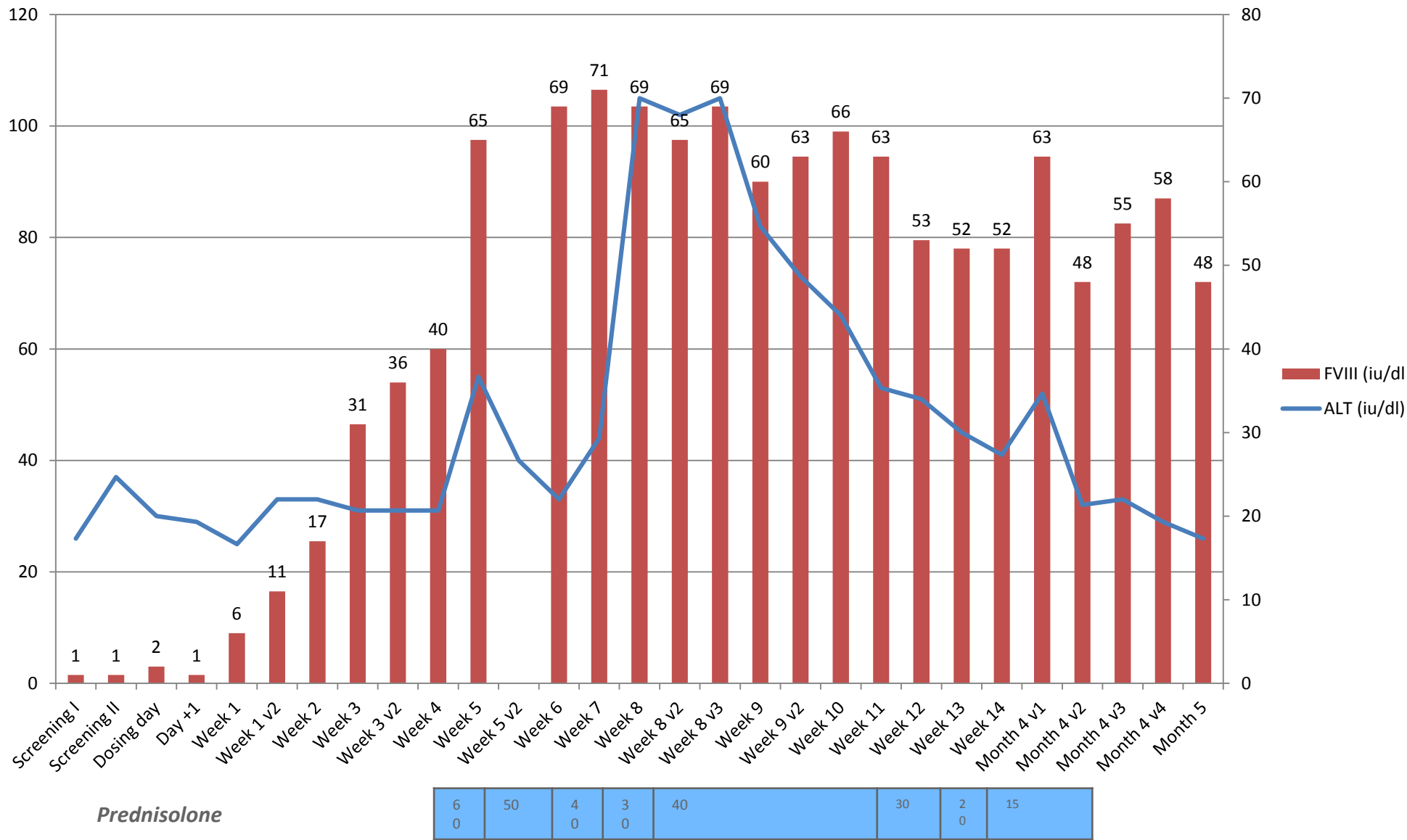
3 Year Phase 2 6e13 vg/kg Data Demonstrates Durable Factor VIII Expression

Rate of FVIII decline continued to be expression level dependent, slowed in year 3, and appears to be approaching plateau

FVIII activity level (IU/dL) time point	Mean (Chromogenic)	Median (Chromogenic)	Mean (One Stage)	Median (One Stage)
Weeks 23-26	68	57	127	100
Week 52	64	60	104	89
Week 104	36	26	59	46
Week 156	33	20	52	30



GO8-4R



Durability: Hemophilia Gene Clinical Trials

<u>Factor IX</u>	NCT	Vector/Gene	AAV-IR	Expression	Durability
▪ UCL/St. Jude	NCT00979238	AAV2/8-LP1-hFIX _{co}	+ 5/10	2-8%	up to 8 years
▪ Shire	NCT01687608	AAV8-sclX	+ 2/7	0-25%	-
▪ Spark	NCT02484092	AAV8(sp _{k100})-hFIX	+	29-35%	up to 2 years
▪ UniQure	NCT02396342	AAV5-hFIX	+ 1/5	5.2-6.9%	up to 2 years
▪ Dimension	NCT02618915	AAVrh10-FIX		10-20%	up to 52 week s
▪ Sangamo	NCT02695160	AAV2/6-ZFN			
<u>Factor VIII</u>					
▪ BioMarin	NCT02576795	AAV5-hFVIII	+	4-300%	up to 2 years - falling
▪ Spark	NCT03003533	AAV8(sp _{k200})-hFVIII			
▪ UCL/St. Jude	NCT03001830	AAV2/8-HLP-FVIII-V3			
▪ Sangamo	NCT03061201	AAV2/6-hFVIII			

Known Unknowns in Gene Therapy

- Consequences of integrating DNA into chromosomal genome?
- How to manage hepatocyte toxicity?
- Where else apart from liver does vector genome go?
- How to overcome existing immunity to AAV?
- How long will transgenic synthesis of factor last?
- Can we repeat treatment with another AAV serotype or viral type?
- Should we treat children despite their growing livers?
- What will it cost?

Where are we now?

We are learning how to do gene therapy for haemophilia one patient at a time

Questions.